

Optimization of novel combi-molecules: Identification of balanced and mixed bcr-abl/DNA targeting properties

Zakaria Rachid,^{b,†} Athanasia Katsoulas,^{b,†} Christopher Williams,^a Anne-Laure Larroque,^b James McNamee^b and Bertrand J. Jean-Claude^{b,*}

^aChemical Computing Group Inc., 1010 Sherbrooke Street West, Suite 910, Montreal, Que., Canada H3A 2R7

^bCancer Drug Research Laboratory, Division of Medical Oncology, Department of Medicine, McGill University/Royal Victoria Hospital, 687 Pine Avenue West Rm. M-719, Montreal, Que., Canada H3A 1A1

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Abstract—Steps toward the identification of combi-molecules with strong abl tyrosine kinase (TK) inhibitory property and significant DNA damaging potential are described. The optimized combi-molecule **13a** was shown to induce approximately twofold stronger abl TK inhibitory activity than GleevecTM and high levels of DNA damage in chronic myelogenous leukemic cells.
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The combi-targeting concept is a novel approach to drug design that seeks to synthesize compounds termed ‘combi-molecules’ capable of binding to kinases involved in cell signaling pathways while remaining DNA reactive.^{1–10} This principle was developed on the premise that cancer cells are endowed with multiple mechanisms to overcome cytotoxic lesions. By inducing a tandem DNA damage and blockade of kinases that control signaling pathways related to the repair of the lesions in the cells, it was expected that the latter could be less prone to evade the death pathways. The chronic myelogenous leukemia (CML) is one of the diseases in which one such tyrosine kinase bcr-abl is known to be involved in two major cytoprotective events: (a) induction or activation of DNA repair proteins (e.g., Rad51),^{11,12} (b) activation of the anti-apoptotic PI3K/AKT pathway.^{13,14} Thus, we surmised that combi-molecules designed to block bcr-abl would on one hand alleviate anti-apoptotic signaling and on the other inflict cytotoxic DNA lesions: a strategy designed to induce enhanced cell-killing in CML cells.

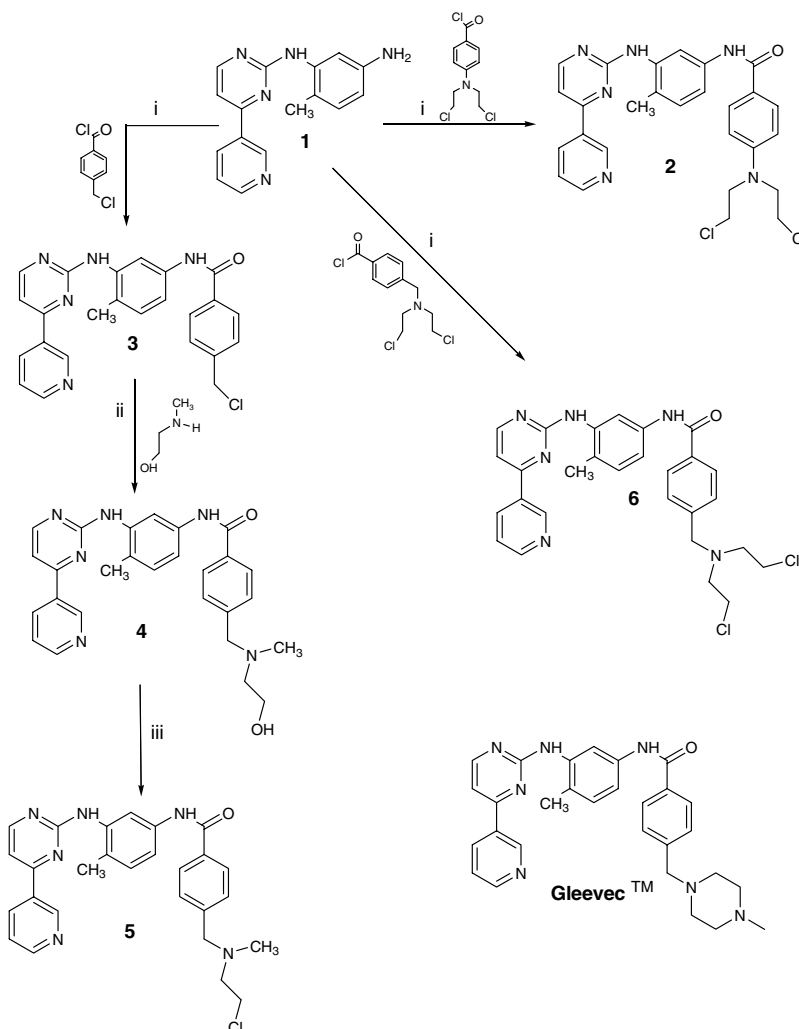
Recently we reported on the synthesis of the first models to demonstrate the feasibility of the approach by designing two types of molecules: (1) those requiring hydrolytic cleavage to generate the DNA alkylating moiety termed ‘type I combi-molecules’,^{8–10} (2) those that can damage DNA without requirement for hydrolytic cleavage (type II). Unfortunately, the first probes synthesized to demonstrate the feasibility of the approach showed disproportionate bcr-abl-DNA targeting properties. The compound **2** (Scheme 1), a type II combi-molecule, while being a potent bcr-abl TK inhibitor (although less potent than GleevecTM) was a weak DNA damaging compound.⁵ The compound **13e** (Scheme 2), a type I combi-molecule, while capable of inducing high levels of DNA damage was a moderate bcr-abl TK inhibitor.^{4,7} Thus, to demonstrate the advantages of this novel approach, a structure optimized to show strong bcr-abl inhibitory potency and fierce DNA damaging potential was urgently needed. Here we report on the steps toward the identification of one such structure **13a**, a triazene with balanced and enhanced bcr-abl-DNA targeting properties.

We first attempted to improve the DNA damaging potential of the type II combi-molecule **2** (because of its superior abl TK inhibitory activity) by inserting a CH₂ group between its mustard moiety and the benzamide function (see **6**, Scheme 1). This is inspired by the fact that the electron density at the nitrogen of **2** is depleted by the benzene ring, thereby reducing its alkylating

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* Corresponding author. Tel.: +1 514 934 1934x35841; fax: +1 514 843 1475; e-mail: bertrandj.jean-claude@mcgill.ca

† These authors contributed equally to this work.



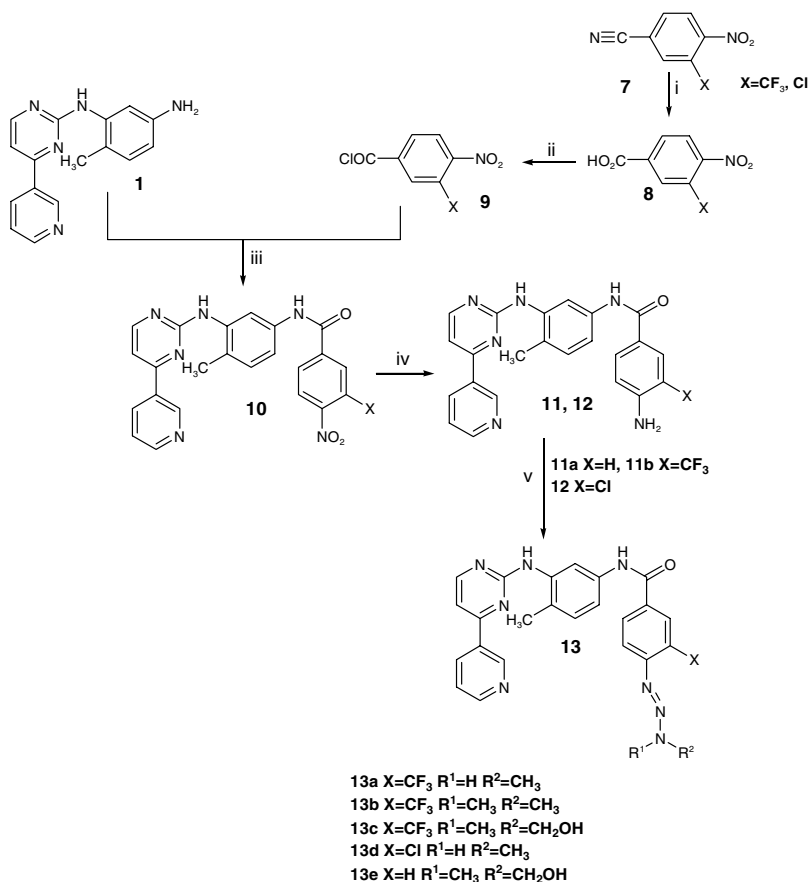
Scheme 1. Reagents and conditions: (i) THF/DIPEA (2 equiv)/0 °C; (ii) DMF/KI (1.1 equiv) at room temperature; (iii) POCl₃/heat.

power. Thus, we surmised that inserting a CH₂ group in this part of the molecule would lead to a compound with not only a strong alkylating power, but also because its structure straddles that of Gleevec™, a strong bcr-abl inhibitory property. The synthesis of compounds **2**, **5**, and **6** proceeded according to [Scheme 1](#). Briefly, amine **1**, synthesized as previously described,¹⁵ was treated with *p*-{*N,N*-bis(2-chloroethyl)amino}benzoyl chloride¹⁶ or *p*-{*N,N*-bis[(2-chloroethyl)aminomethyl]}benzoyl chloride¹⁷ to give compounds **2** or **6**. Treatment of **1** with 4-chloromethylbenzoyl chloride gave **3** which was stirred in DMF with *N*-methylethanolamine to provide **4**. Chlorination of amino-alcohol **4** with POCl₃ gave the hemi-mustard **5** in good yield. Compound **5** was a poor bcr-abl TK inhibitor and inactive against K562 cell. Likewise, compound **6** was more than 50-fold less potent than Gleevec™ and virtually inactive against the CML cell line K562 ([Table 1](#)).

Molecular modeling was used to account for the failure of compound **6**, the design of which was simply based on structural overlap with Gleevec™. As depicted in [Figure 1](#), the phenyl ring of compound **2** ([Scheme 1](#)) is anchored into a hydrophobic pocket of bcr-abl, allow-

ing its mustard moiety to occupy a region in the receptor where minimum steric clash can occur. In contrast, the CH₂ group of **6** protrudes the mustard moiety into an orientation where its steric bulk can induce clash with the pocket. Reducing the size of the mustard by creating a hemi-mustard **5** ([Scheme 1](#)) resulted in a compound with greater affinity for bcr-abl but with a potency approximately 75-fold less than that of Gleevec™. Thus the type II strategy involving the bulky mustard group was abandoned. However, data acquired from this work allowed us to better understand the mode of binding of the combi-molecules, most particularly the anchorage of the benzamide ring into a narrow hydrophobic pocket and the requirement for a non-bulky DNA damaging tail. For the latter property, the triazene tail already demonstrated to induce high levels of DNA damage in the pyrido-pyrimidine system could now be selected as the group of choice.⁷ However, for enhancing the interaction of the abl recognition moiety, the challenge remained.

Based upon results obtained for **2**, we thought it of interest to enhance hydrophobic interactions of the benzamide moiety with the hydrophobic pocket.



Scheme 2. Reagents and conditions: (i) HCl_{concd}/heat; (ii) SOCl₂, THF/heat; (iii) pyridine(2 equiv)/CH₂Cl₂/0 °C; (iv) Fe/ethanol/AcOH/heat; (v) CH₃CN/NOBF₄/−5 °C/ether then addition of alkylamine correspondent or 1:30 mixture of aqueous methylamine (40%) and aqueous formaldehyde (37%).

Table 1. Cytotoxicity and Abl tyrosine kinase inhibitory activities by different compounds

Compound	Inhibition of abl kinase IC ₅₀ ^a , μM	IC ₅₀ ^b , μM (K562)
Gleevec™	0.04	0.18
2 ⁵	0.22	2.46
5	3.11	79.72
6	>10	>100
11b	0.156	0.316
11a	>10	68
13a	0.028	0.113
13b	0.018	0.052
13c	0.033	0.187
13d	0.180	9.98
13e ⁷	1.057	3.15

^a IC₅₀ values were determined by using the c-abl tyrosine kinase protein. Values are means of two separate experiments.

^b IC₅₀ values were obtained from the MTT assay.

Interestingly, during the course of this development, Asaki et al.,¹⁸ coincidentally addressing the same issue, demonstrated that compounds with small hydrophobic substituents, for example, Cl and CF₃ on the benzamide moiety could show enhanced binding affinity when compared with Gleevec™. Thus inspired by our results on **2** and the latter development, we designed combi-molecules containing: (a) a Cl and CF₃ group on the phenyl

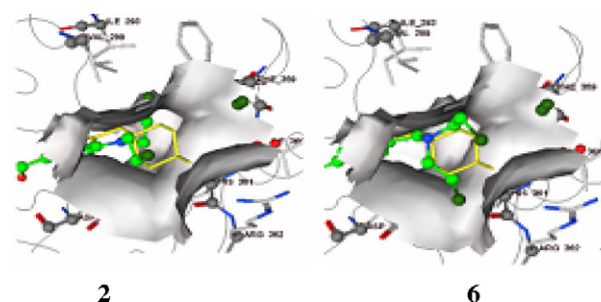


Figure 1. Compounds **2** and **6** in 1IEP (X-ray structure of Gleevec™ shown in yellow).

ring of the benzamide moiety, and (b) a small 1,2,3-triazene tail carrying one or two methyl groups at the 3-position.

The synthesis of compounds **13a–d** proceeded according to an adaptation of a strategy developed for the synthesis of **13e**.⁷ Briefly, commercially available compound **7** was hydrolyzed under acidic condition to give the acid **8** which was chlorinated with SOCl₂ to give the benzoyl chloride **9**. Treatment of **1** with the benzoyl chloride **9** in methylene chloride/pyridine (2 equiv) at 0 °C gave the nitro compounds **10**, which were reduced with Fe in ethanol to provide the amines **11a–b**, and **12**.

Diazotization of these amines with nitrosonium tetrafluoroborate in acetonitrile followed by the addition of the desired amine and neutralization with triethylamine gave triazene conjugates **13a–d**.¹⁹

Interestingly, abl TK enzyme assays showed that compound **13d** containing a 3-Cl group showed potency similar to that of **2** but 10-fold better than **13e**. The combi-molecules with the more hydrophobic CF₃ group, **13a–c**, showed abl TK inhibitory potency not only superior to that of **13e** but also to that of GleevecTM. Compound **13b**, the dimethyl analog of **13a**, was the most potent of the series, which we believe is due to its more hydrophobic dimethyl function. It should be noted that the latter compound could not be considered to be a combi-molecule in vitro since like all dimethyl-triazenes,^{20,21} it cannot be converted to **13a** without metabolic activation.

One of the special features of type I combi-molecules is their ability to not only block their target on their own but also to further degrade into another metabolite with bcr-abl TK inhibitory activity that sustains inhibition of bcr-abl TK. HPLC analysis showed that more than 70% of **13a** was degraded to **11b** as early as 3 h after incubation in cell culture medium at 37 °C. It should be remembered here that the previous non-optimized combi-molecule **13e** degraded to generate the amine **11a** which was found to be virtually inactive against bcr-abl TK-expressing cells.^{4,7} In contrast, compound **11b** is 400-fold more potent than the latter amine and only threefold less potent than GleevecTM. Molecular modeling study suggests that the superior potency of **11b** may be mediated as for the combi-molecule by the ability of the CF₃ group to occupy a hydrophobic subpocket in the ATP binding site of the receptor (Fig. 2). The 1IEP crystal structure of Abl kinase bound to Gleevec²² was used as a starting point to construct compounds **13** analogs in the Abl pocket. The 1IEP PDB file was loaded into the MOE software package²³; all crystallographic water molecules were deleted and hydrogen

atoms were added. The ligand coordinates of GleevecTM from the 1IEP X-ray structure were used as a template to manually construct starting structures for the compounds **13** compounds in the pocket of the 1IEP crystal structure. MMFF94x^{23,24} partial charges were assigned to all the atoms in the system, and each ligand structure was minimized to a gradient of 0.01 kcal/Å mol in the presence of the receptor using the MMFF94x forcefield in MOE. The receptor atoms were held fixed throughout all the minimizations. In all cases, the structures minimized to coordinates very similar to the GleevecTM X-ray coordinates. Furthermore, the minimizations show that there is sufficient room for the CF₃ substituent in the hydrophobic subpocket. Experimental evidence for occupation of this pocket is provided by the 2HIW PDB crystal structure of complexed abl kinase, which shows a CF₃ substituent on the inhibitor occupying this subpocket.²⁵ For the optimized combi-molecule **13a**, Poisson–Boltzmann electrostatic maps of the entire abl pocket²⁶ (Fig. 3) show a 3 kcal/mol hydrophobic isocontour in the subpocket, suggesting that hydrophobic contact with this region will yield an approximate 3 kcal/mol

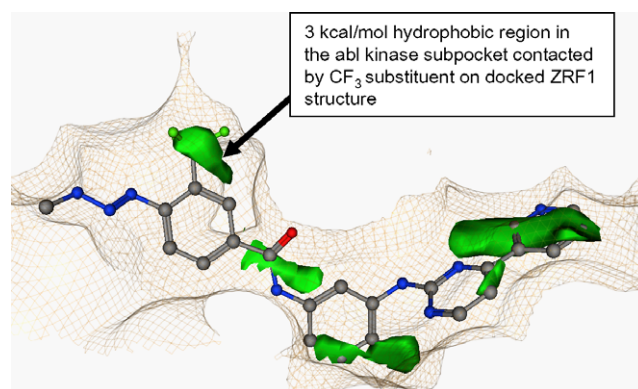


Figure 3. Compound **13a** docked in 1IEP abl pocket. Green surfaces reflect the 3 kcal/mol electrostatic energy isocontour for hydrophobic contacts within the pocket.

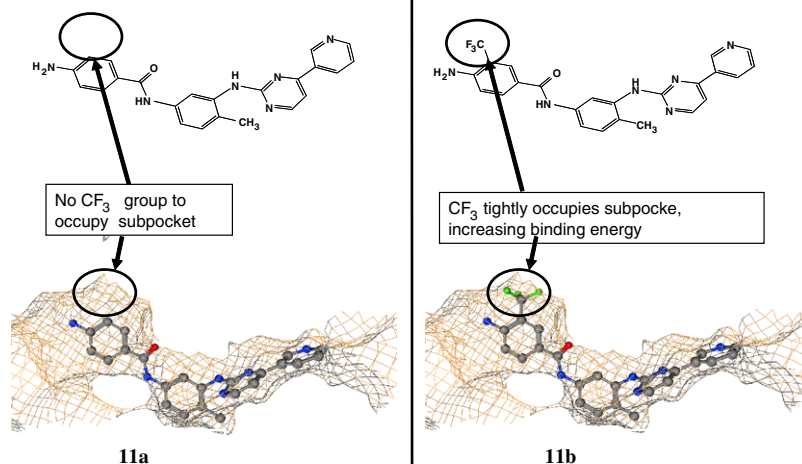


Figure 2. Occupation of 1IEP subpocket by ortho CF₃ group of **11b**.

gain in ligand binding energy. Indeed, this predicted 3 kcal/mol binding energy gain correlates well (Table 2) with the increase in affinity observed between pairs of compounds that differ only by CF₃ substituent [**11a**/**11b** and **13e**/**13c**].

In order to determine the translation of the now proportionate binary targeting property of the combi-molecules, we studied their potency against the bcr-abl expressing CML cells K562. The results showed that **13a** is twofold more potent than GleevecTM and fivefold more potent than **11b**. Furthermore, its DNA damaging potential was demonstrated by the comet assay. Interestingly, compound **13a** induced significant levels of DNA damage in the Mo7p210 transfected cells (Fig. 4).

This study conclusively demonstrated that the potency of the combi-molecules could be refined by enhancing the hydrophobicity of the benzamide triazene carrier with a binding mode similar to that of GleevecTM. The now augmented abl TK inhibitory potency (activity superior to GleevecTM) has conferred to these Type I combi-molecules of the triazene class, strong and balanced bcr-abl/DNA targeting properties. To our knowledge, this is the first report on such a combi-molecule with balanced bcr-abl/DNA targeting properties. A complete study on its biological mechanism of action (blockade of abl phosphorylation in whole cells, analysis of the DNA damage response pathways p53, p21, induction of apoptosis (Bax), and comparative analysis with individual combinations of bcr-abl + DNA damaging agents will be reported elsewhere.

Table 2. Binding energy gains from –CF₃ substitution on benzamide moiety in GleevecTM analogs

Compound pair	Inhibition of abl kinase IC ₅₀ , μM		ΔΔG ^a (kcal/mol)
	X = –H	X = CF ₃	
11a / 11b	67	0.156	3.6
13e / 13c	1.1	0.033	2.1

^a Binding energy gains estimated from experimental IC₅₀ values using $\Delta\Delta G = -rt \ln(IC_{50CF_3}/IC_{50H})^{27}$.

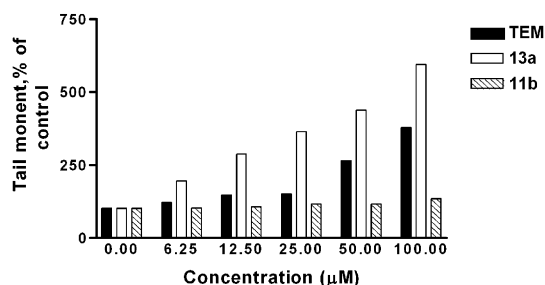


Figure 4. Quantitation of DNA damage using the alkaline comet assay. DNA damage induced by **13a**, **11b** and Temozolomide (TEM) in the M07/p210 cell line. Tail moment was used as a parameter for the detection of DNA damage in M07/p210 cells exposed to the drugs 30 min.

Acknowledgments

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- Experimental data for **13a**. 1-{N-[4-Methyl-3-(4-pyridin-3-yl-pyrimidin-2-ylamino)-phenyl]-3-trifluoromethyl-benzamide}-methyl triazene. The amine **11b** (100 mg, 0.22 mmol) was dissolved in dry acetonitrile (10 mL) and after cooling to –5 °C, a solution of nitrosonium tetrafluoroborate (50 mg, 0.5 mmol) suspended in acetonitrile was added. The clear solution was stirred for 1 h at –5 °C and a mixture of ether (10 mL), water (2 mL), triethylamine (0.43 mL) and aqueous methylamine (40%) (0.2 mL) was added dropwise. The mixture was subsequently stirred at 0 °C for 15 min, the reaction was extracted with ethyl acetate dried then evaporated and the precipitate that formed triturated and washed with

- ether to give **13a** (50 mg, 42%) as a pure solid. ESI m/z 529.0 (MNa)⁺; ¹H NMR (400 MHz, DMSO-*d*₆) δ 11.30 (br s, 1H, N=N–NMeH), 10.33 (s, 1H, NH), 9.27 (s, 1H, ArH), 8.99 (s, 1H, ArH), 8.67 (d, 1H, $J = 4.8$ Hz, ArH), 8.51–8.46 (m, 2H, ArH), 8.26 (s, 1H, ArH), 8.18 (d, 1H, $J = 12.4$ Hz, ArH), 8.06 (s, 1H, ArH), 7.64 (d, 1H, $J = 11.2$, $J = 11.2$ Hz, ArH), 7.53–7.42 (m, 3H, ArH), 7.21 (d, 1H, $J = 11.2$, ArH), 3.07 (s, 3H, NHCH₃), 2.22 (s, 3H, CH₃); ¹³C NMR (75 MHz, DMSO-*d*₆) 168.7, 164.5, 162.3, 161.9, 160.2, 152.1, 151.8, 148.9, 138.5, 138.4, 137.7, 135.1, 133.2, 132.9, 131.4, 130.8, 128.5, 126.6, 124.5, 118.1, 118.0, 117.6, 108.2, 31.4, 18.3.
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